

# Candida Biofilms

## Perspectives from a Clinician

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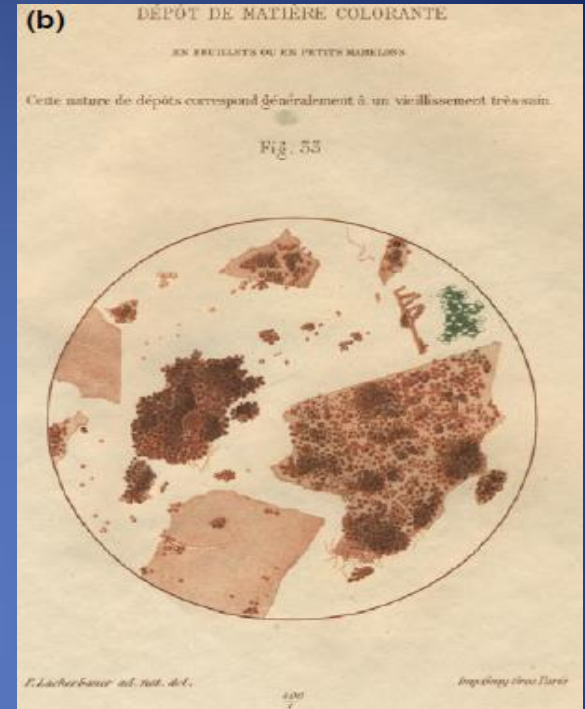
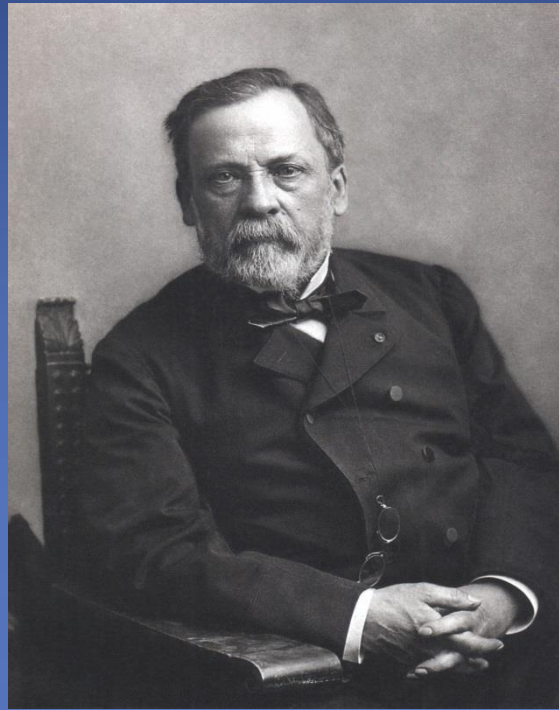
# **BIOFILM VS. BIOFILM INFECTIONS**

# Bio-film

- Referring to bacterial adhesion, aggregation and multiplication on surfaces, was used in marine microbiology to distinguish adhering (**sessile**) bacteria from free swimming (**planktonic**) bacteria as early as 1933

# Luis Pasteur

(1822-1895)

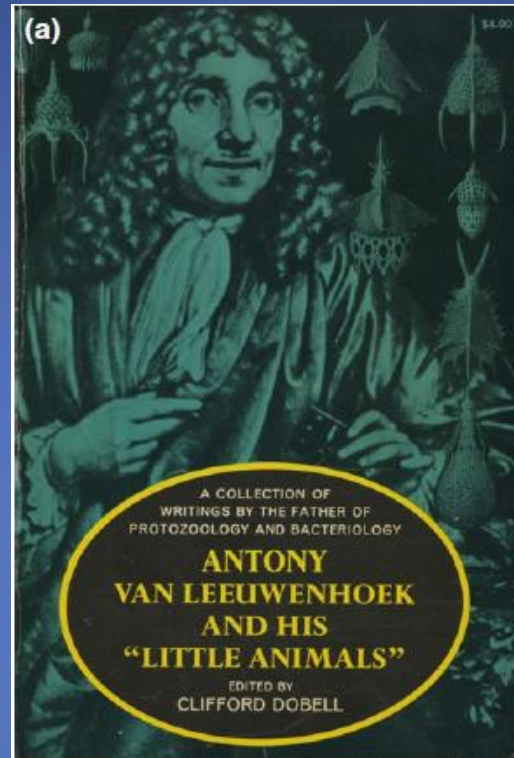


An aggregate of microbial cells adherent to a living or nonliving surface, embedded within a matrix of extracellular polymeric substances of microbial origin.

Høiby N *Pathog Dis* 2014

Flemming HC et al. *Nat Rev Microbiol* 2010

# Antony van Leeuwenhoek (1632-1723)



Aggregated, microbial cells surrounded by a polymeric self-produced matrix, which may contain host components.

Høiby N *Pathog Dis* 2014

Hall-Stoodley L et al. *FEMS Immunol Med Microbiol* 2012

# Driving Forces for Biofilm Formation



# Biofilm in Medicine

- 1970-1972, cystic fibrosis patients with chronic *Pseudomonas aeruginosa* lung infection.

Høiby N et al. *Acta Pathol Microbiol Scand B* 1973

Høiby N *Acta Pathol Microbiol Scand B* 1974

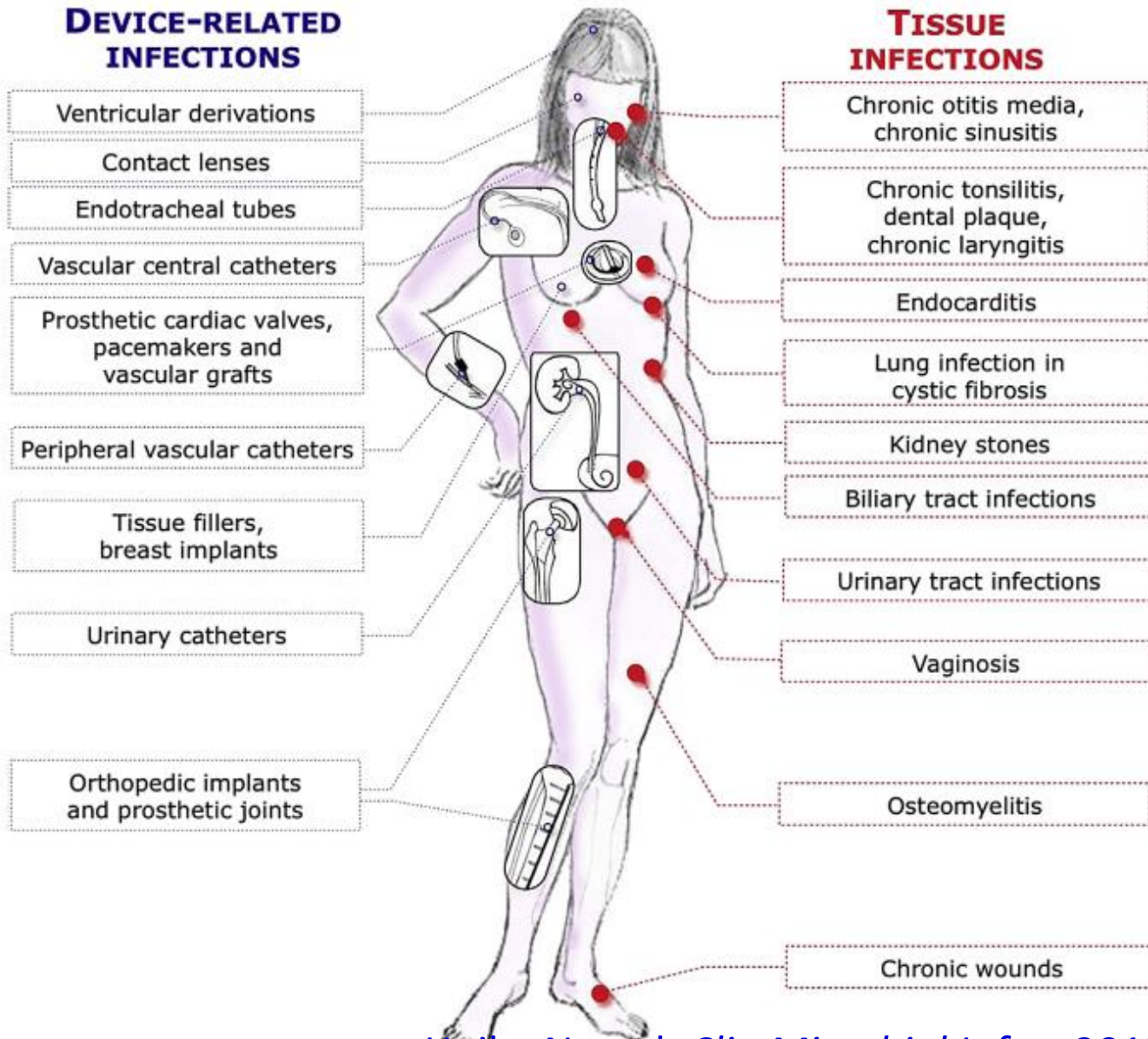
- 1978-1980, *Candida*-induced denture stomatitis

Budtz-Jørgensen *J Am Dent Assoc* 1978

Theilade J et al. *J Biol Buccale* 1980



# Biofilm Associated Infections





# *Candida* infections of Medical Devices

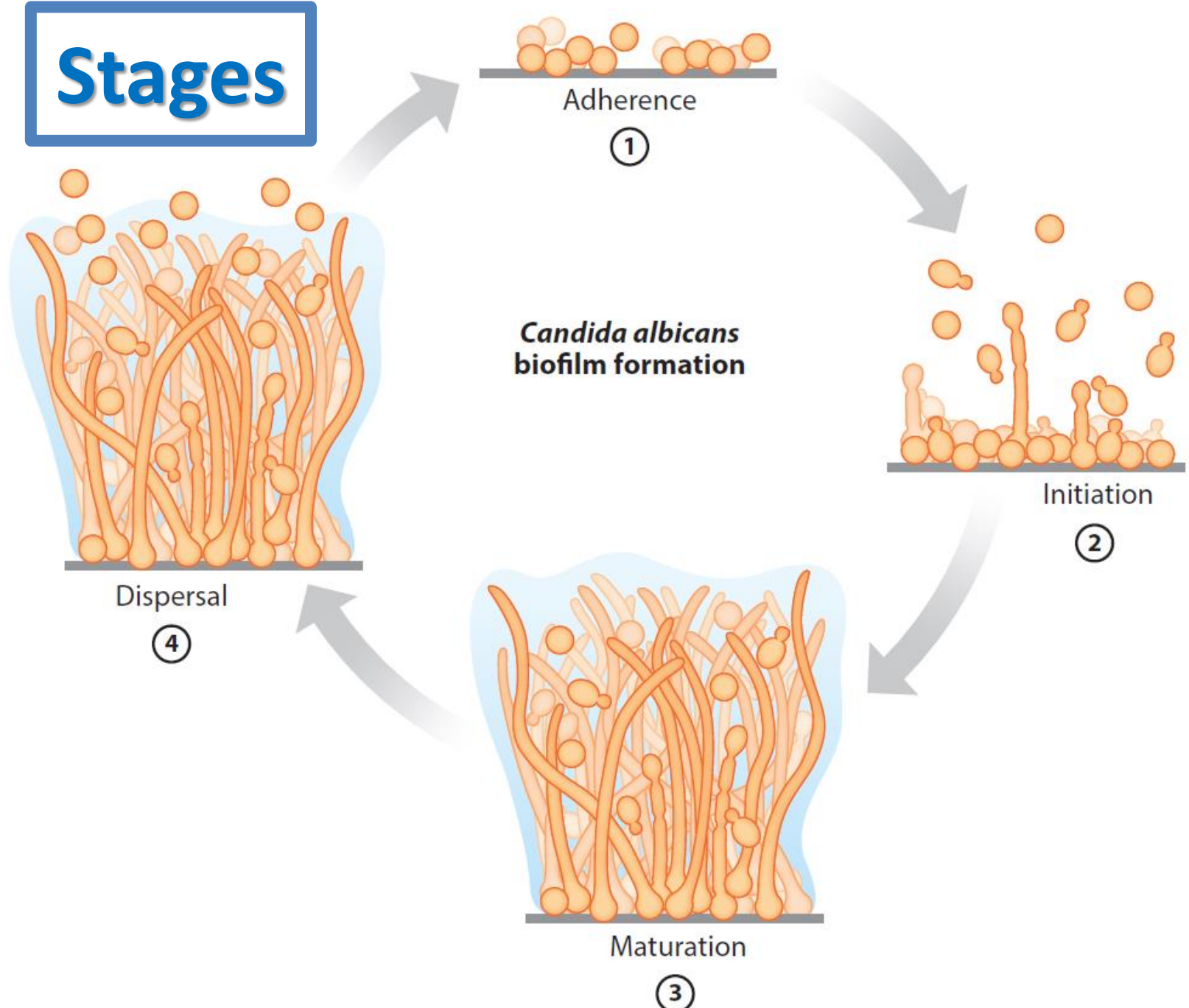
Device	Infection rate (%)	Proportion of <i>Candida</i> spp. Infection (%)	Risk factor
<b>Vascular catheters</b>	3-8	10	Similar with risk factors of candidemia
<b>Prosthetic valves</b>	2.9	2-10	Prior bacterial IE; Prolonged antibiotics use; IV cath; IVDU
<b>Pacemakers</b>	0.5-7	4.5	Nil.
<b>VP shunt</b>	6-15	1	Prior or concurrent meningitis; Broad spectrum antibacterials; Bowel perforation; abdomen surgery
<b>PD catheters</b>	23	2.4-7	Recent bacterial peritonitis; Prior antibacterials; SLE
<b>Joint prostheses</b>	1-3	<1	Nil.

\* Removal needed to achieve cure for all medical devices

Cauda R et al. *Drug*, 2009

# ***BASICS OF CANDIDA* BIOFILM**

# Stages



# How to Quantify Biofilms?

## Crystal violet stain

- Staining the metabolically active and inactive cells in mature biofilms
- **Biomass** production

## XTT reduction assay

- XTT [2,3-bis(2-methoxy-4-nitro-5-sulfo-phenyl)-2*H*-tetrazolium-5-carboxanilide]
- Yellow salt that is reduced by dehydrogenases of **metabolically active** cells to a colored formazan product

Measured colorimetrically with a microtiter plate reader

Taff HT, et al. *Med Mycol* 2012

# Factors affecting *Candida* Biofilm *in vitro*

1. Fluid flow shear
2. Substrate
3. Nutrients
4. *Candida* species and strains
5. Microbial cohabitants

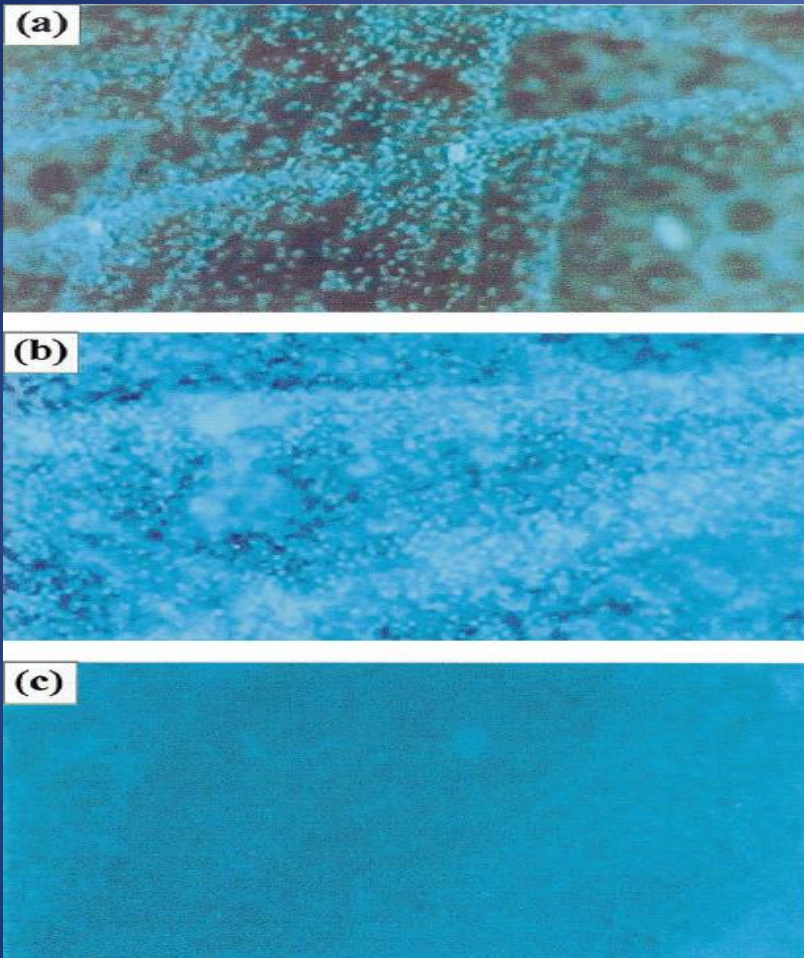
# Comparisons of Visualization Methods

## Biofilm Morphology & Architecture

	Fluorescence microscopy	Scanning electron microscopy (SEM)	Confocal scanning laser microscopy (CSLM)
Advantages	Quick method as a screening tool	Surface topography of biofilms at very high magnification	three-dimensional reconstruction of undisturbed biofilm
Disadvantages	Low magnification	Dehydration artifacts in the biofilm matrix	

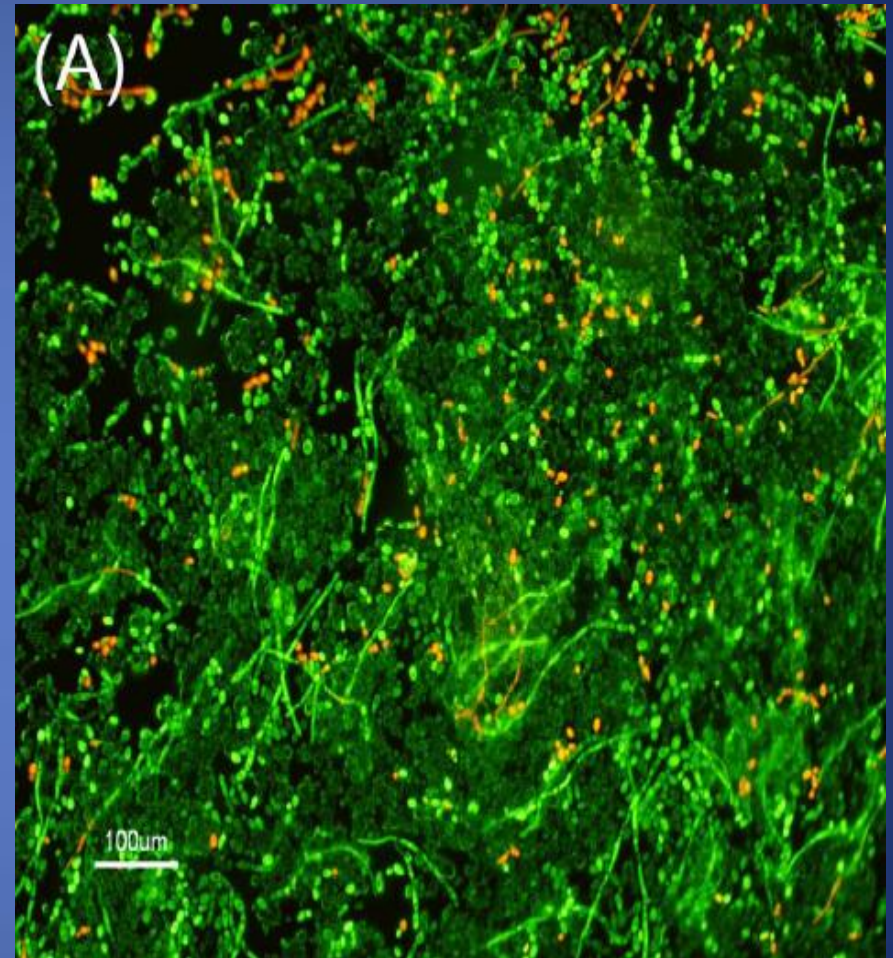


# Fluorescence microscopy



Calcofluor-White

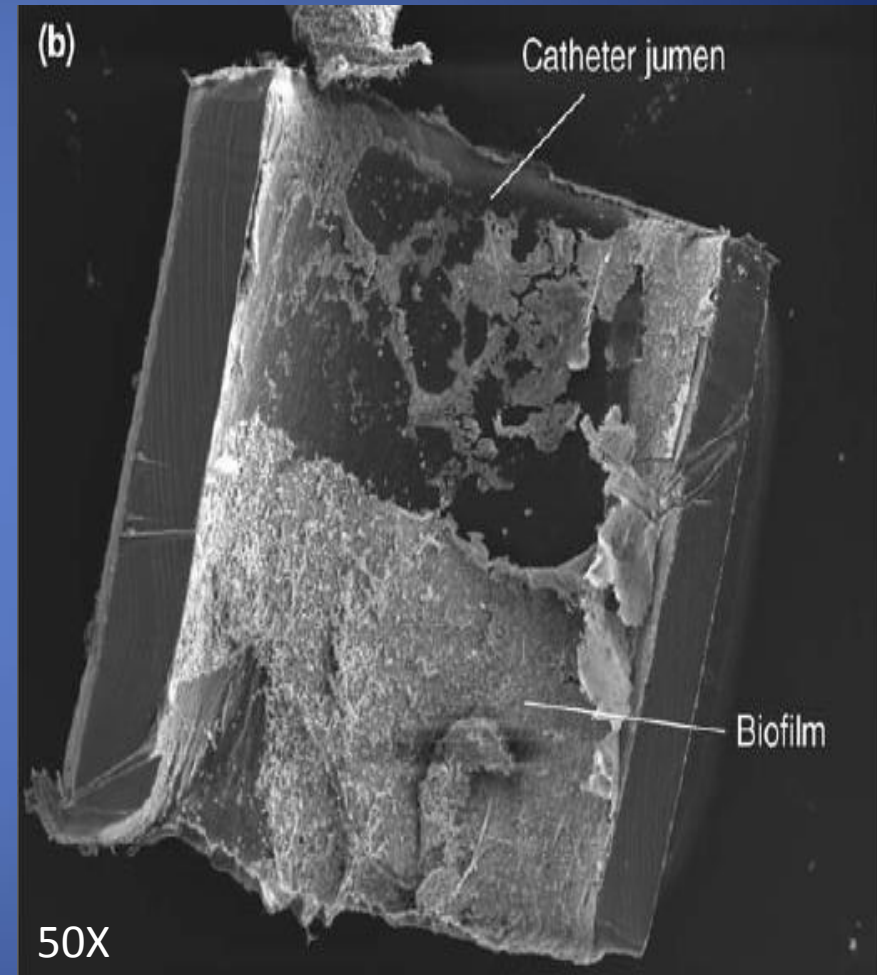
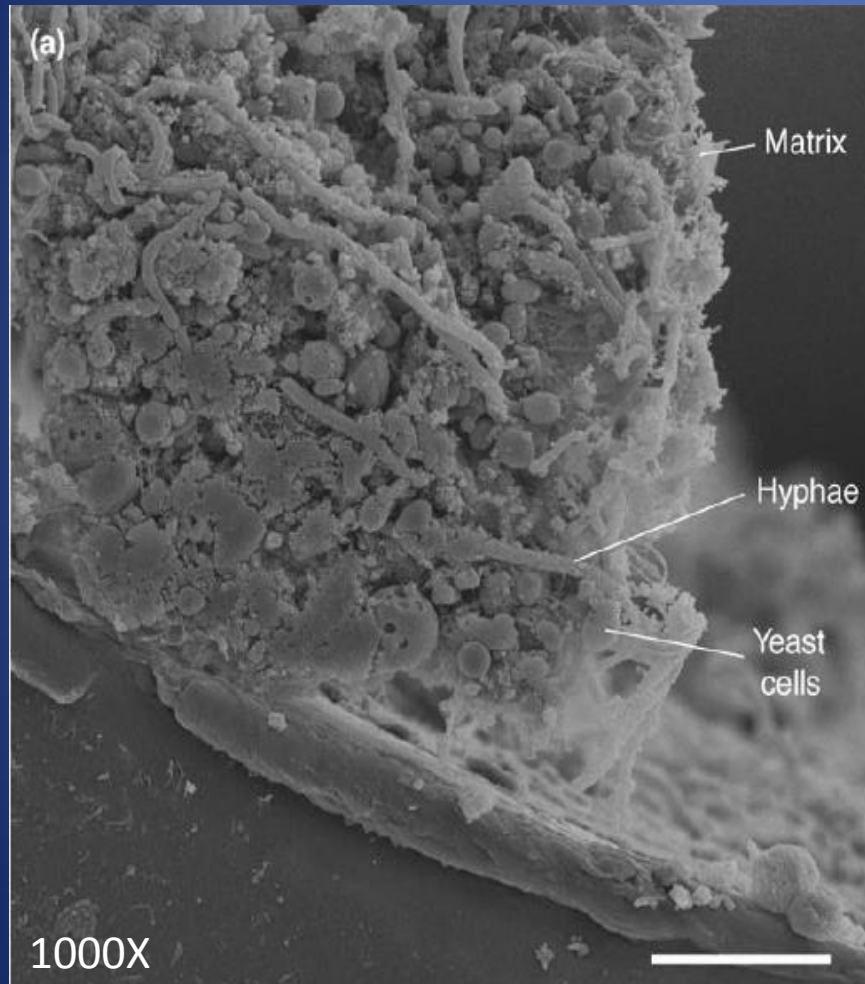
Chandra J et al. *J Bacteriol* 2001



SYTO 9 dye/ Propidium iodide

Tøndervik A et al. *PLoS One* 2014

# Scanning Electron Microscopy



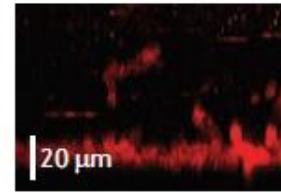


# Confocal Scanning Laser Microscopy

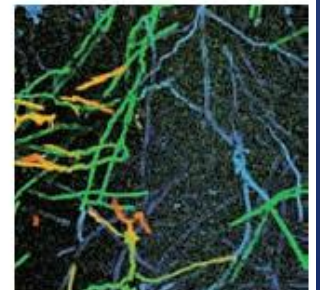
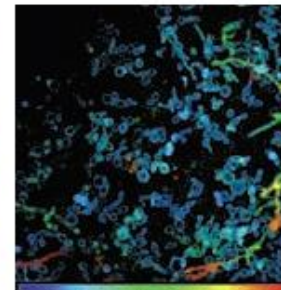
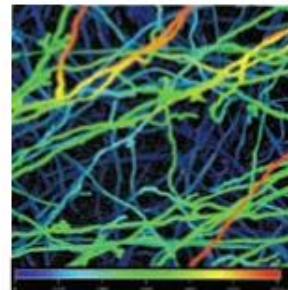
## Concanavalin A-Alexa Fluor

Finkel J et al. *Nat Rev Microbiol*  
2011

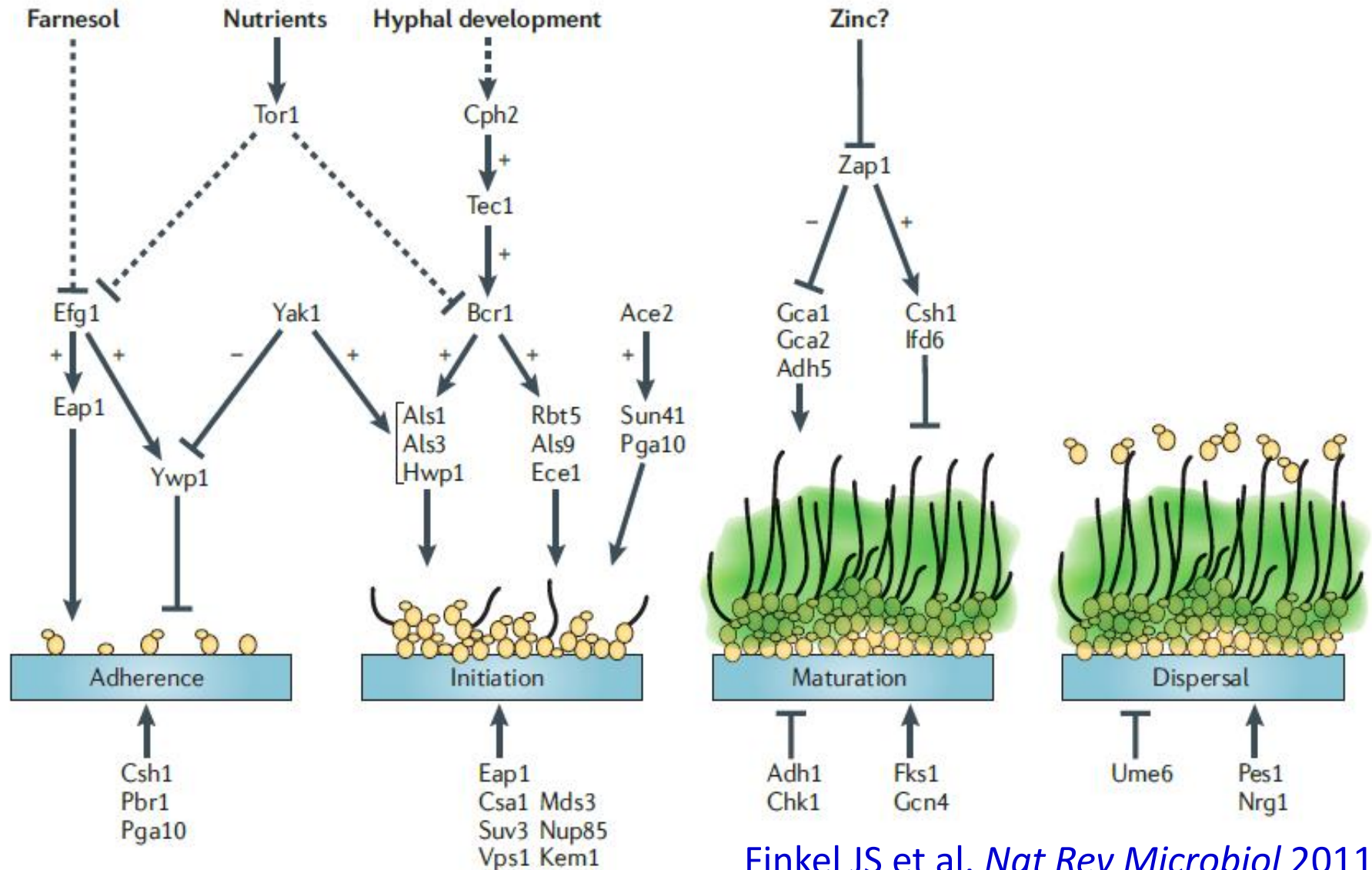
CSLM  
side view



CSLM  
depth view



# Proteins that Function in Biofilm Development



Finkel JS et al. *Nat Rev Microbiol* 2011

# Selected Genes in Biofilm Development

Molecular function of gene products*	Role of gene product	Genes
Transcription factors	Positive	<i>ACE2<sup>‡</sup>, BCR1, CPH1, CZF1<sup>‡</sup>, EFG1<sup>‡</sup>, FLO8<sup>‡</sup>, GCN4, TEC1<sup>‡</sup>, UME6<sup>‡</sup> and NRG1<sup>‡</sup></i>
	Negative	<i>ZAP1</i>
Cell wall-related proteins	Positive	<i>ALS1, ALS2<sup>‡</sup>, ALS3, ALS4, ALS5, ALS7, ALS9, CSA1, EAP1, FKS1, HWP1, HWP2, OCH1, PGA1, PGA10<sup>‡</sup>, PMT1<sup>‡</sup>, PMT2<sup>‡</sup>, PMT4, PMT6, RBT1, RBT5 and SUN41<sup>‡</sup></i>
	Negative	<i>YWP1</i>
Alcohol dehydrogenases	Positive	<i>ADH5</i>
	Negative	<i>ADH1, CSH1 and IFD6</i>
Protein kinases	Positive	<i>CBK1<sup>‡</sup>, GIN4<sup>‡</sup>, IRE1<sup>‡</sup>, MKC1 and YAK1<sup>‡</sup></i>
	Negative	<i>CHK1 and TOR1,</i>
Drug efflux pumps	Positive	<i>CDR1, CDR2 and MDR1</i>
Glucoamylases	Positive	<i>GCA1 and GCA2</i>
Other functions <sup>§</sup>	Positive	<i>CAT2, ECE1, KEM1<sup>‡</sup>, MDS3<sup>‡</sup>, NDH51, NUP85<sup>‡</sup>, PBR1, PES1, PDX1, RIX7, SUV3<sup>‡</sup>, VAM3<sup>‡</sup> and VPS1<sup>‡</sup></i>



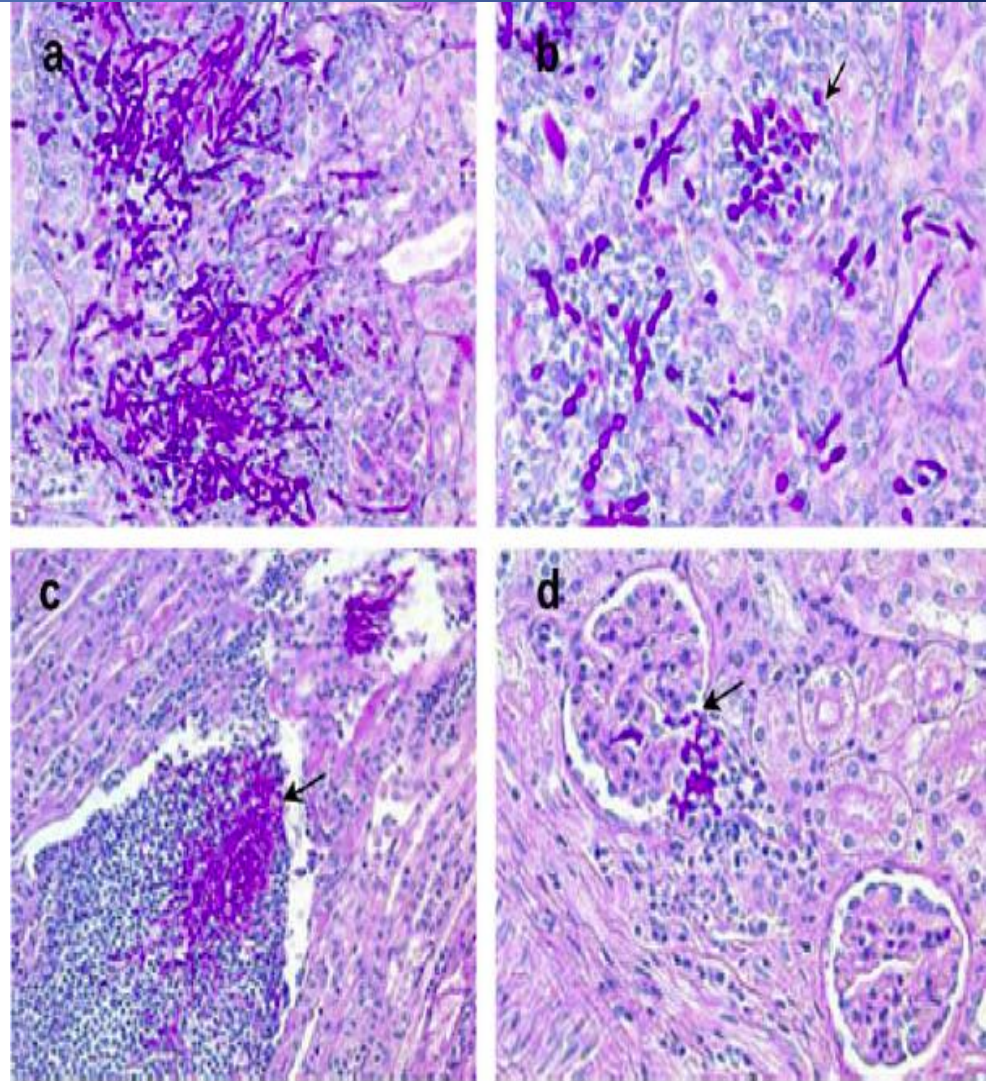
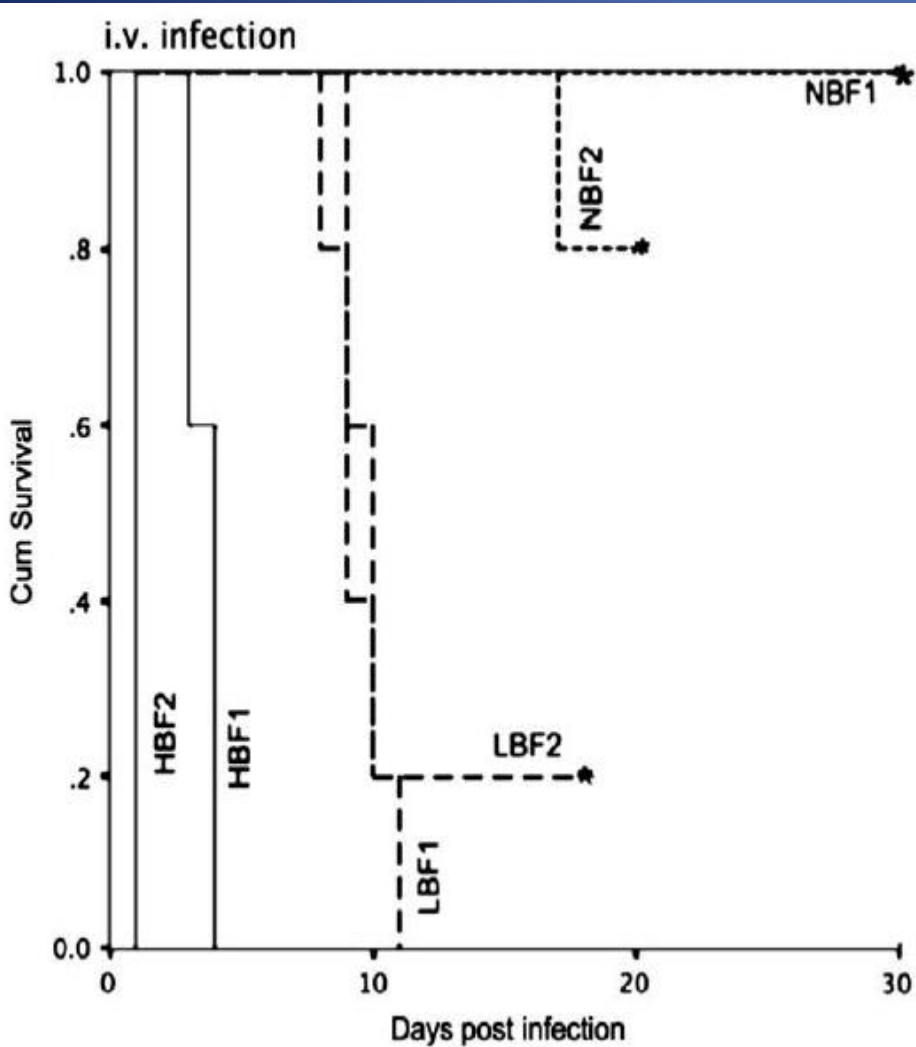
A regulator of filamentation

Finkel JS et al. *Nat Rev Microbiol* 2011

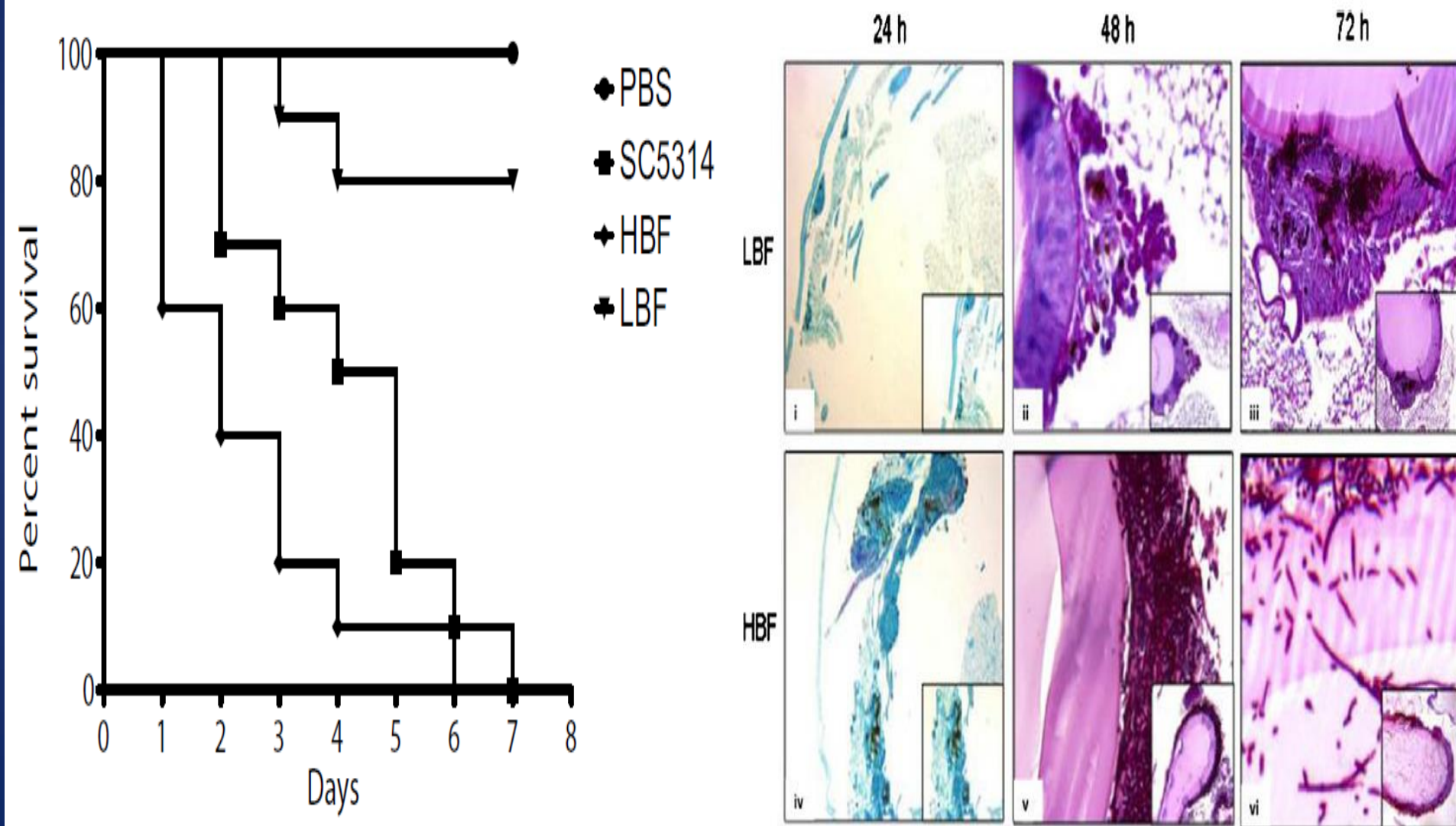
# BIOFILM PATHOGENESIS IN *CANDIDA* INFECTION



# Virulence in Intravenous Murine Infection Model



# Virulence in *Galleria mellonella* Infection Model



# Planktonic vs. Sessile, Azole

Species	No. of isolates tested	Type of MIC <sup>a</sup>	No. of isolates for which indicated MIC (μg/ml) was:												
			0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	32	64	>1,024
<i>C. albicans</i>	12	MIC for planktonic cells			3	4	1	3			1				
		MIC <sub>50</sub> for sessile cells									1	1	2	1	7
		MIC <sub>80</sub> for sessile cells													12
<i>C. parapsilosis</i>	12	MIC for planktonic cells				1	4	4	3						
		MIC <sub>50</sub> for sessile cells											2		10
		MIC <sub>80</sub> for sessile cells													12
<i>C. tropicalis</i>	10	MIC for planktonic cells			1	6	1	2							
		MIC <sub>50</sub> for sessile cells									2				8
		MIC <sub>80</sub> for sessile cells													10
<i>C. glabrata</i>	9	MIC for planktonic cells								1	2	5	1		
		MIC <sub>50</sub> for sessile cells									1				8
		MIC <sub>80</sub> for sessile cells													9

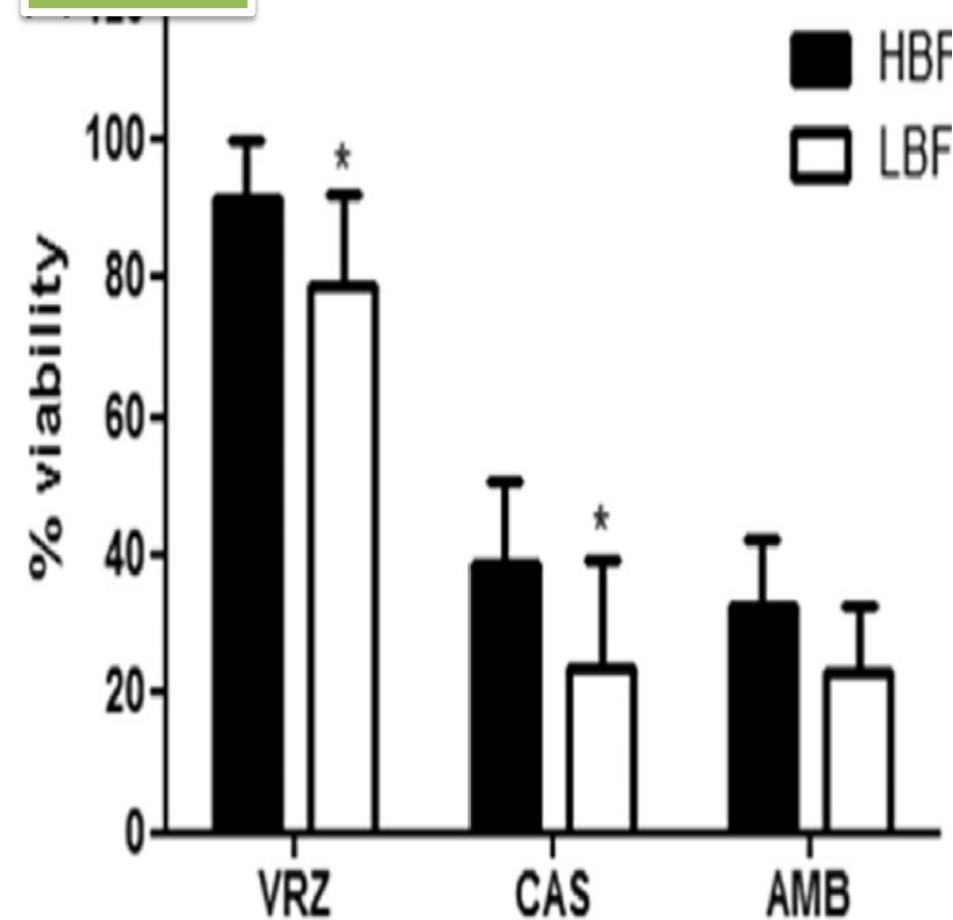


# Planktonic vs. Sessile, Candin

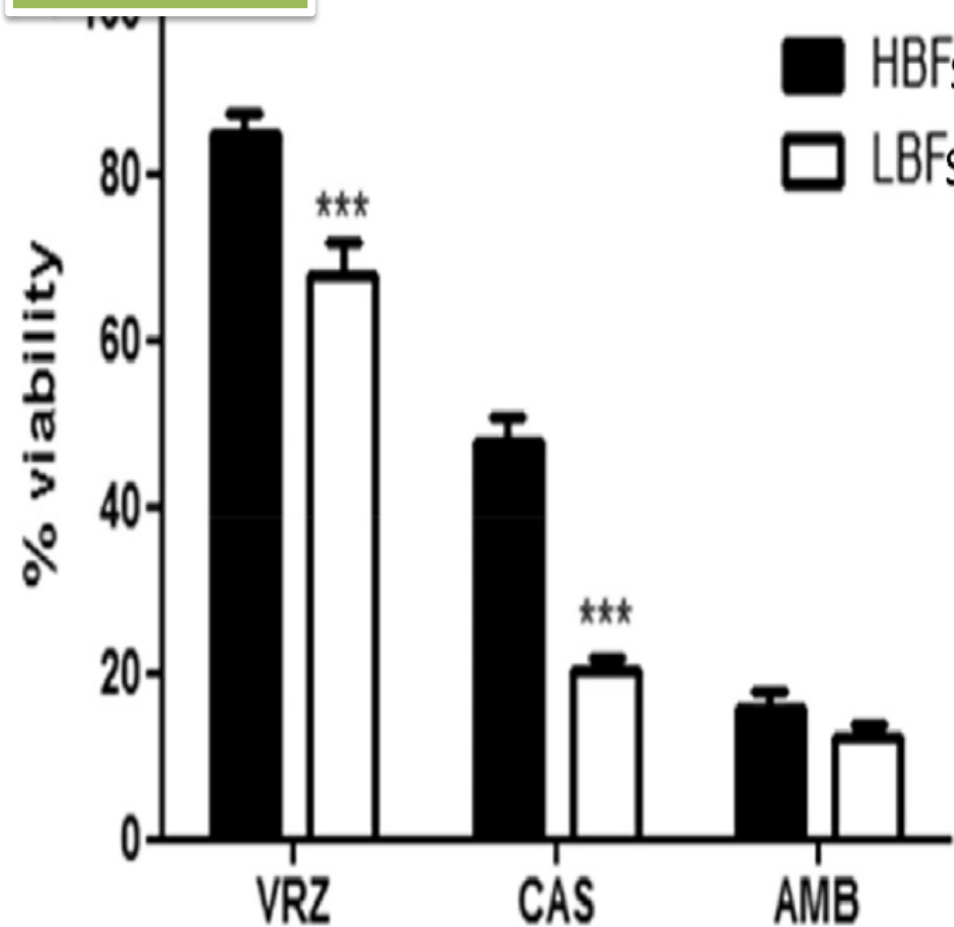
Drug	Species	No. of isolates tested	Type of MIC <sup>a</sup>	No. of isolates for which indicated MIC (μg/ml) was:										
				0.03	0.06	0.12	0.25	0.5	1	2	4	8	16	>16
Caspofungin	<i>C. albicans</i>	12	MIC for planktonic cells	6	4	2								
			MIC <sub>50</sub> for sessile cells		1	1	1	9						
			MIC <sub>80</sub> for sessile cells					9	3					
	<i>C. parapsilosis</i>	12	MIC for planktonic cells				6	5	1					
			MIC <sub>50</sub> for sessile cells						1	2	2	2		5
			MIC <sub>80</sub> for sessile cells											12
	<i>C. tropicalis</i>	10	MIC for planktonic cells	4	5	1								
			MIC <sub>50</sub> for sessile cells						6	2				2
			MIC <sub>80</sub> for sessile cells											10
	<i>C. glabrata</i>	9	MIC for planktonic cells	2	5	2								
			MIC <sub>50</sub> for sessile cells			1	1	6	1					
			MIC <sub>80</sub> for sessile cells					3	6					

# High vs. Low Biofilm Formation, Antifungal

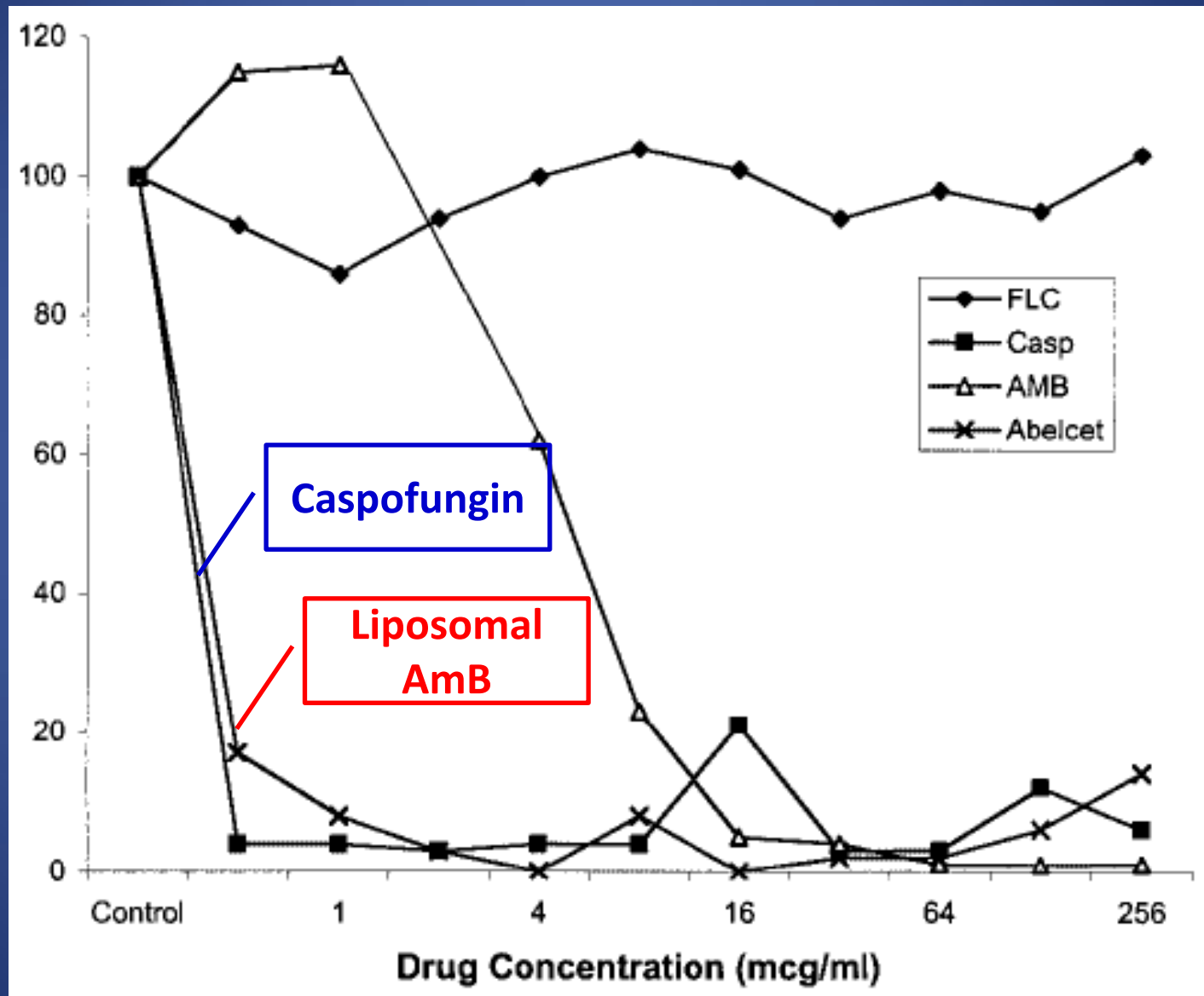
2 mg/L



200 mg/L



# Candins and Liposomal AmB Are Better!



Kuhn DM et al. *Antimicrob Agents Chemother* 2002



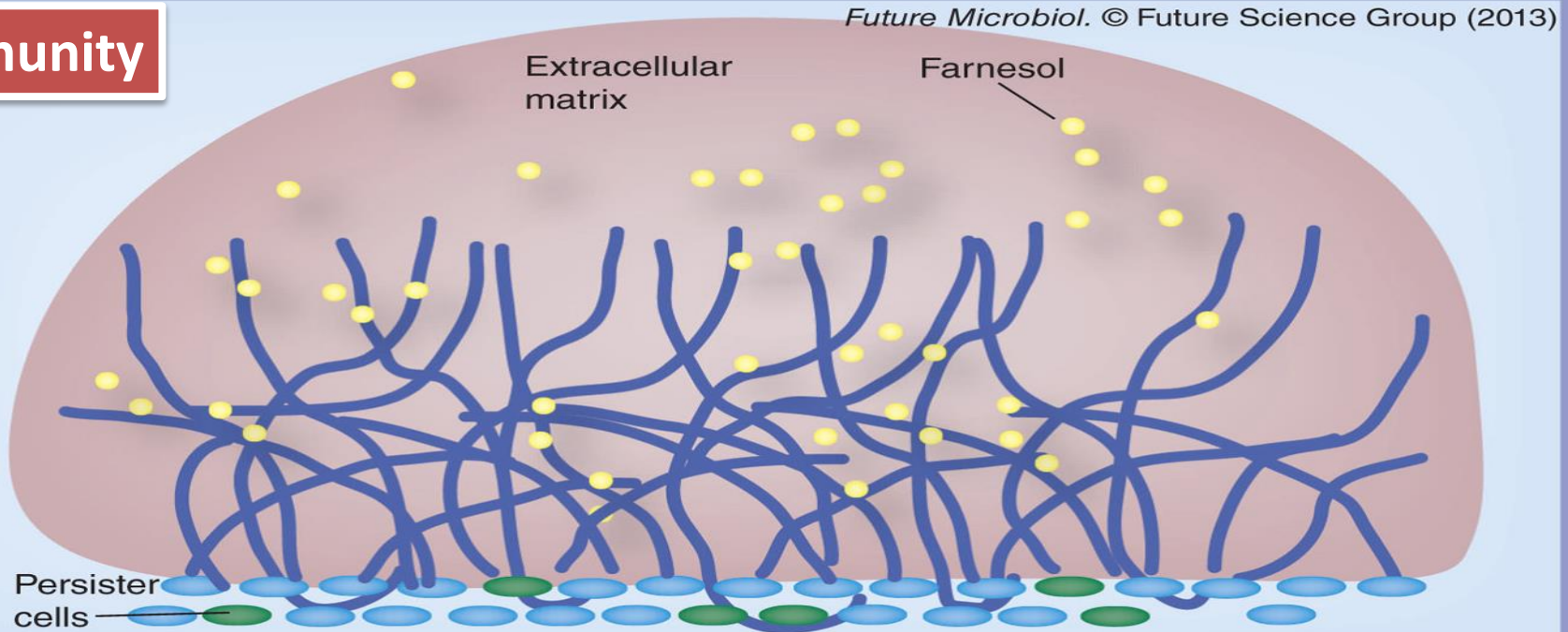
# Biofilms Resistance

## □ Possible mechanisms

1. Alteration in membrane sterol composition **(Early)**
2. Overexpression of drug efflux pump during early phase of biofilm formation **(Intermediate & Mature)**
3. Extra-cellular matrix retards the diffusion of drugs across biofilm, especially in a mixed-species biofilm

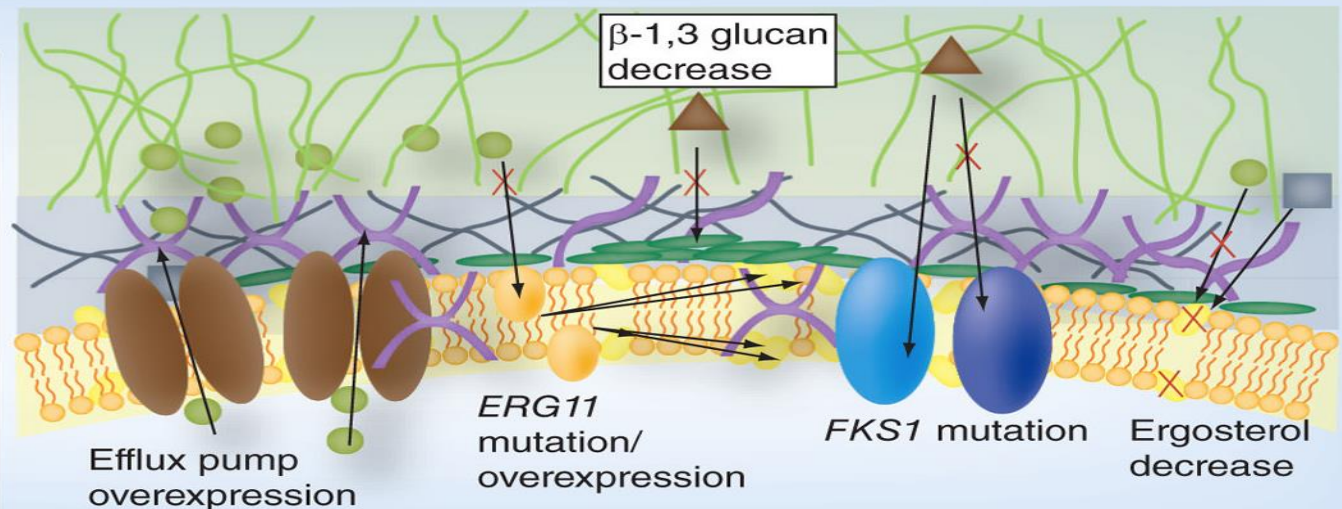
# Candida Biofilm Resistance Mechanisms

## Community



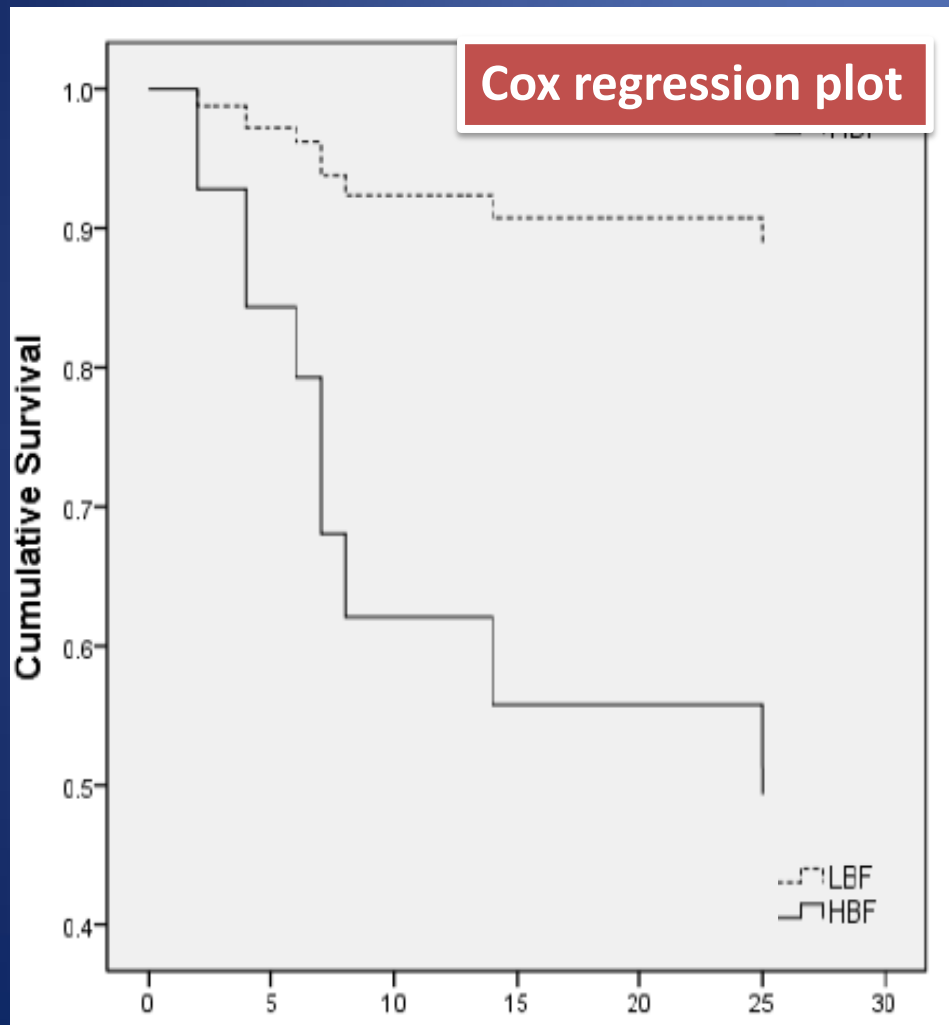
## Cell

- Fks1p
- Plasma membrane
- Ergosterol
- $\beta$ -1,3 glucan
- Unknown matrix components
- Nonglucan cell wall
- Chitin
- Efflux pumps
- Azole
- Amphotericin B
- Echinocandin
- ERG11

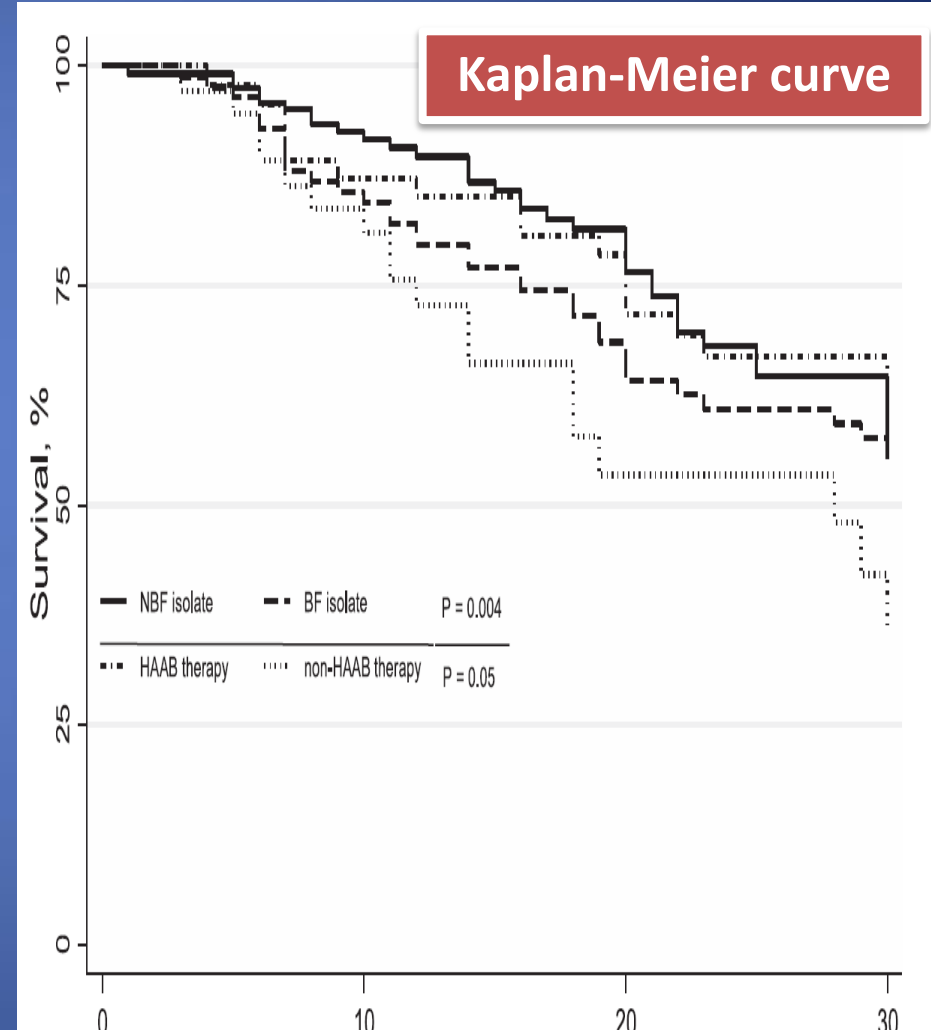


# **CLINICAL RELEVANCE & MANAGEMENT**

# Impact of Biofilm Formation & Antifungal on Patient Survival



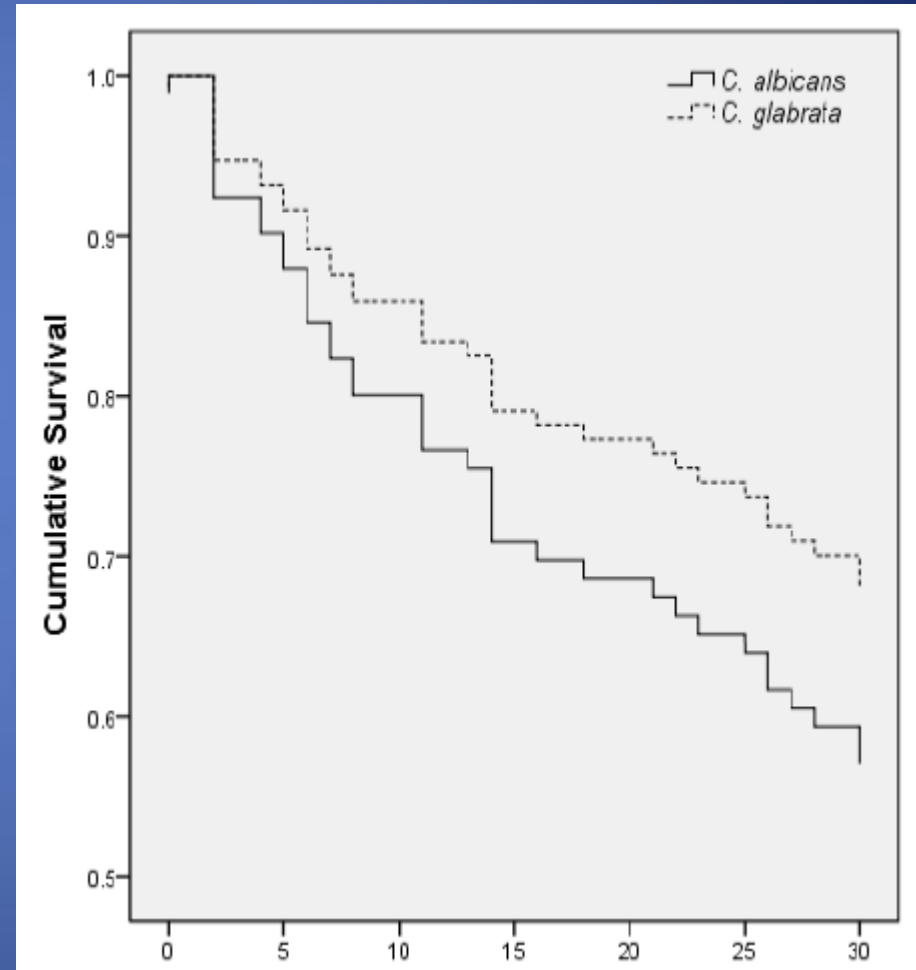
Rajendran R et al. *Clin Microbiol Infect* 2016

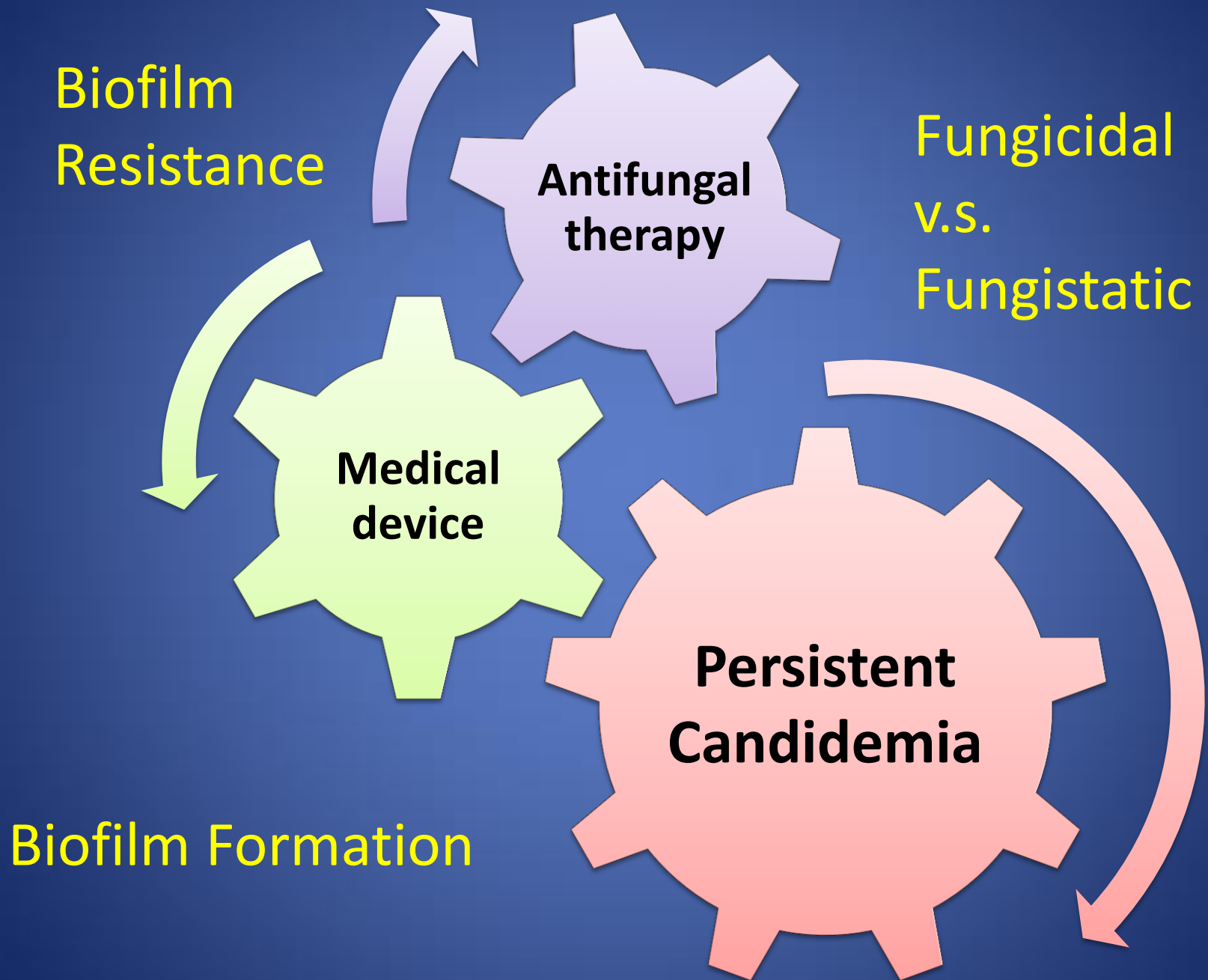


Tumbarello M et al. *PLoS One* 2012

# Species Difference, mainly on *C. albicans*

<i>Candida</i> species	OR (95% CI)	<i>p</i> <sup>a</sup>
<i>C. albicans</i>	3.90 (1.72–8.83)	<0.001
<i>C. parapsilosis</i>	4.16 (1.46–11.82)	0.003
<i>C. tropicalis</i>	0.88 (0.54–1.45)	0.62
<i>C. glabrata</i>	1.46 (0.32–6.62)	0.61
Other <sup>b</sup>		0.34
Total	2.76 (1.55–5.00)	<0.001







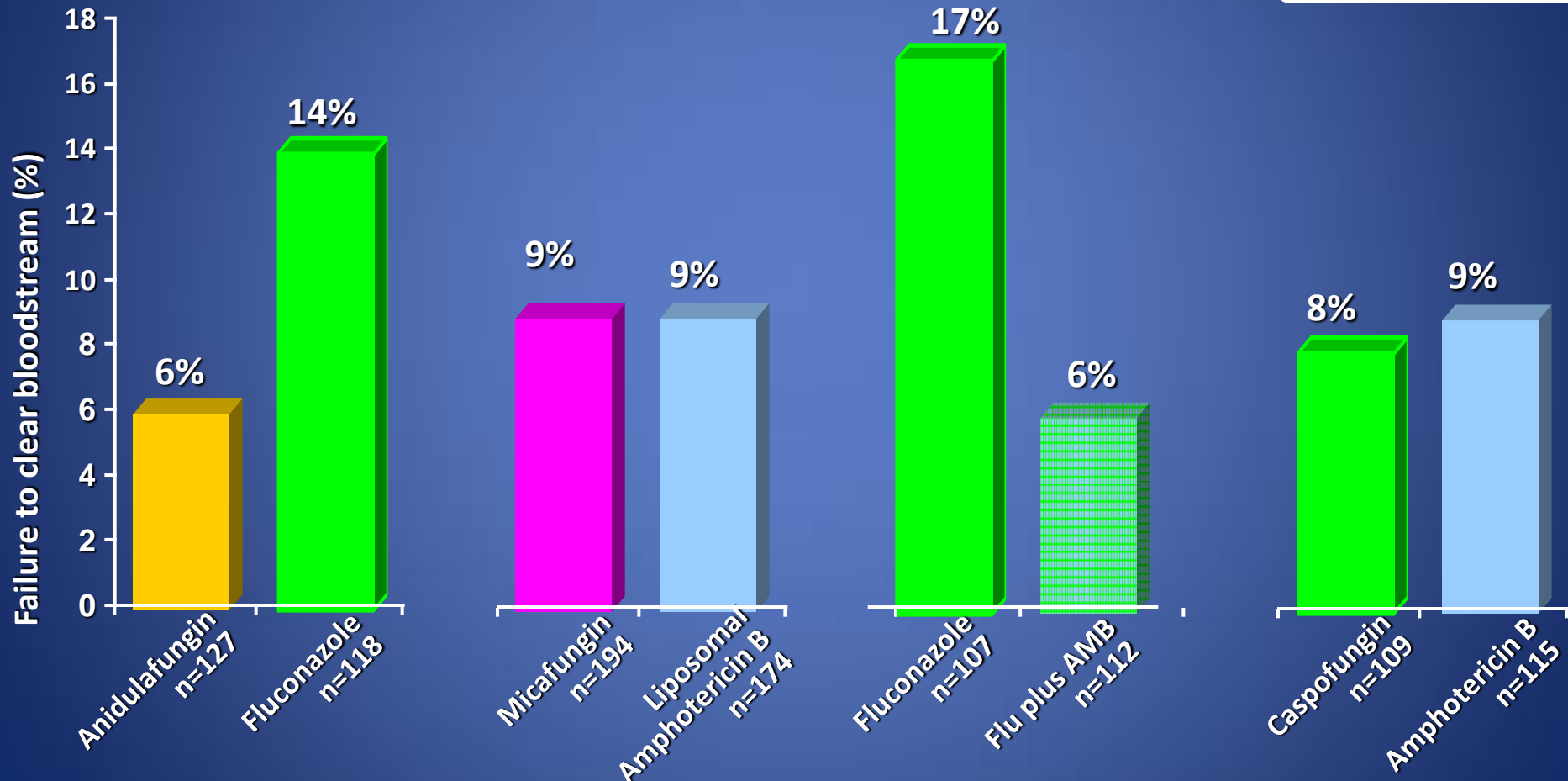
# In published studies, end-of-treatment fungal persistence rates have ranged from 6% to 17%

Reboli et al, 2007

Kuse et al, 2007

Rex et al, 2003

Mora-Duarte et al,  
2002



Reboli AC et al. *N Engl J Med*. 2007;356:2472-2482.  
Mora-Duarte J et al. *N Engl J Med*. 2002;347:2020-2029.

Kuse ER et al. *Lancet Infect Dis*. 2007;369:1519-1527.  
Rex JH et al. *Clin Infect Dis*. 2003;36:1221-1228.

# Etiologies of Persistence

## 1. Intravascular infection

- ❑ Endocarditis
- ❑ Suppurative thrombophlebitis

## 2. Metastatic sites

- ❑ Osteoarticular infection
- ❑ Endophthalmitis

## 3. Inserted medical device

- ❑ Intravascular catheter (CVC, Hickman, etc.)
- ❑ Prosthetic valves
- ❑ Joint protheses
- ❑ Pacemaker

## 4. Pharmacology

- ❑ Adequate dosing
- ❑ Drug resistance

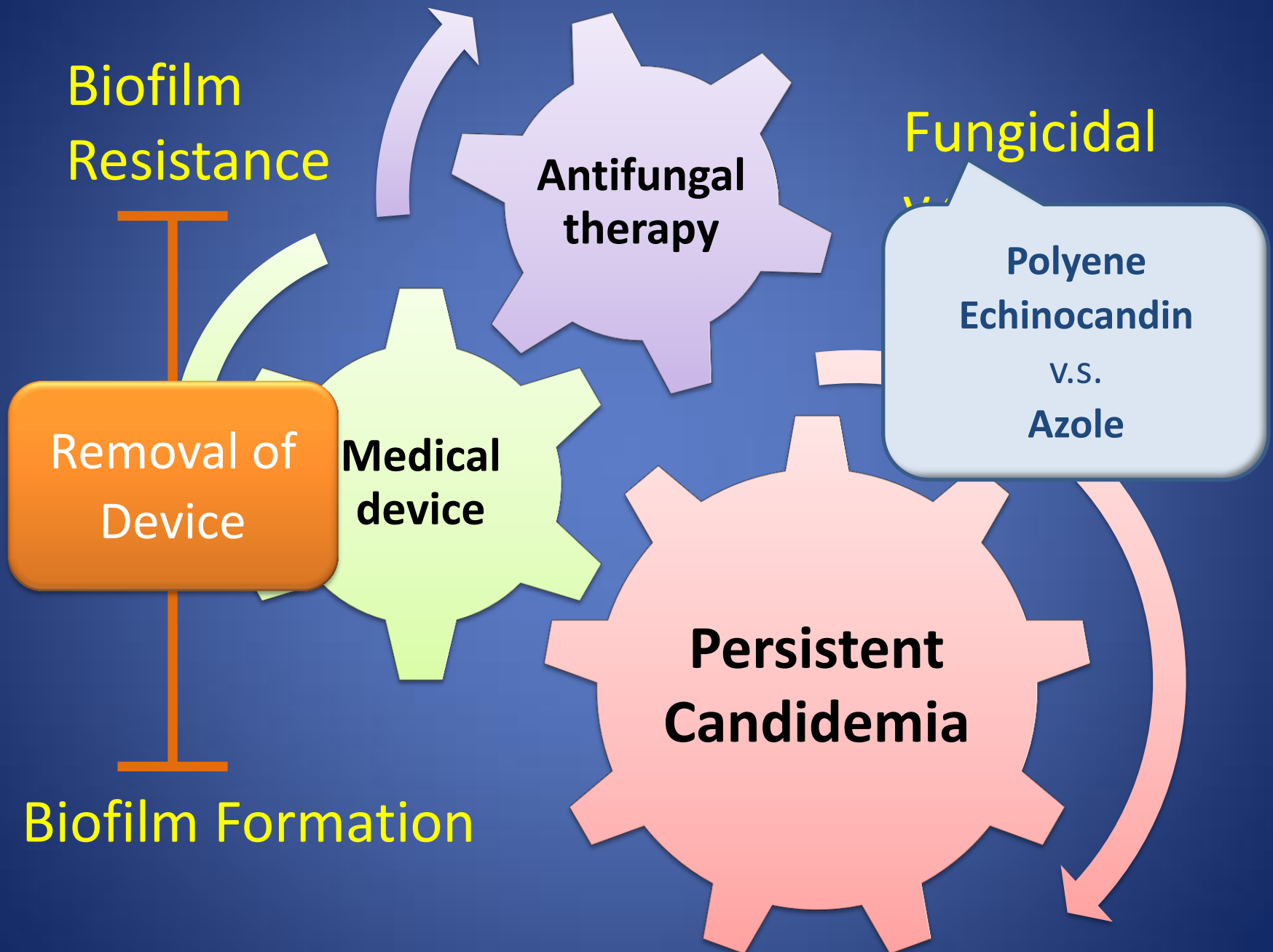
## 5. Host immunity



**Biofilm Formation**

# How to Deal with Persistent Candidemia?

- ❑ Removal of intravascular devices if possible
- ❑ Finding out the possibility of other metastatic infection sites
- ❑ The susceptibility testing of the pathogen
- ❑ Change antifungal agents
  - Azoles → **polyenes or candins**
- ❑ Ameliorating the immunosuppression status





**Thanks! Any Comments?**